

### **REMARKS**

Claims 41-75, and 84-97 have been cancelled. Claims 1, 7, 9, and 14 have been amended. Claims 17-40 are withdrawn from consideration. Rejoinder of claims 17-40 is respectfully requested. Claim 7 is amended to correct a minor typographical error. Support for the remaining amendments is found in the existing claims and the specification as discussed below. Accordingly, the amendments do not constitute the addition of new matter.

The amendments are fully supported by the specification e.g., paragraph 186 of the published application (present specification, paragraph 0168) and Example 3, paragraph 277 of the published application (present specification, paragraph 240). Thus, no new matter has been added. Applicant respectfully requests the entry of the amendments and reconsideration of the application in view of the amendments and the following remarks.

#### **Rejections under 35 U.S.C. § 112**

Claims 98-109 are rejected under 35 U.S.C. § 112 as being allegedly non-enabled. The Examiner alleges that claims 98-109 are non-enabling because "the specification, while being enabling for encapsulation of pancreatic cells, does not reasonably provide enablement for all claimed cells". Further, the Examiner states that the specification in paragraph 48, describes densities for pancreatic cells only. Applicants respectfully traverse the rejection as follows.

The Examiner asserts that Applicants only describe encapsulation of "pancreatic cells" and not any other type of cell. Applicants submit that the specification provides support for encapsulating other than pancreatic cells (Examples 1-11 of the specification). For example, Example 12 and corresponding FIGs. 28- 30, describe viability post-encapsulation of mouse insulinoma cell line [FIG.28; paragraph 312], a monkey kidney cell line [FIG. 29A; paragraph 313]; and cell aggregates produced from primary liver cells (hepatocytes) from both human and mouse origin [FIG.30; paragraph 314]. All cells were successfully coated and viable post-encapsulation based on fluorescent light with FDA/EB staining. Additionally, Examples 15-20 of the specification, describe encapsulation of human or animal fibroblasts, vascular cells, or various non-tumorigenic cell lines, and genetically engineered cell lines for encapsulation (paragraphs 328-333 of the specification).

Examples 1-11 of the specification, clearly describe the encapsulation of pancreatic islets. The skilled artisan based on these descriptions can apply these teachings to other cell types

without undue experimentation. Examples 12-15 of the specification show that by using substantially the same methods as that described for encapsulation of pancreatic cells, Applicants were able to encapsulate lung (Figure 29) and primary liver cells (Figure 30), and further demonstrated that these cell types remained viable at least up to 2 weeks post-encapsulation. Therefore, it is clear that cells other than pancreatic cells are encompassed and envisioned by Applicants and that the disclosure is not limited to pancreatic cells. Other cell types are described and enabled.

The Examiner also states that the specification in paragraph 48, describes densities for pancreatic cells only. Applicants respectfully submit that the Examiner appears to be contradictory in this and the obviousness rejection below. Because at the same time the Examiner alleges that Applicants only describe densities for pancreatic cells and not other cell types, in the obviousness rejection below, the Examiner alleges that cell densities per se are within the skill of one in the art. Thus, if cell densities are allegedly “within the skill of one in the art” then Applicants written description of pancreatic cell densities is more than sufficient to provide teachings for other cell types.

However, independent of the obviousness rejection, from Examples 12-14 and Tables 3-6 of the specification, it is clear that Applicants clearly describe how to determine the “curative dose” of cells which should and can be encapsulated. Tables 3-6 lay out the parameters and provide guidance to the skilled artisan. Thus, determining cell densities of other cell types requires some experimentation but not undue experimentation.

Therefore, Applicants respectfully request that the rejections of claims 98-109 based on lack of enablement be withdrawn.

**Rejections under 35 U.S.C. § 103(a)**

Claims 1-16, 76-83 and 98-109 are rejected under 35 U.S.C. §103(a) as being obviousness over WO 00/53159 (WO'159). Applicants respectfully traverse this rejection as follows.

The Examiner alleges that the claims are obvious over WO'159 because WO'159 describes the identical encapsulation coating except for the cell densities, which the Examiner alleges is within the skill of one in the art.

Claims 1, 9, and 14 have been amended to recite that the cell aggregates are conformally coated, wherein the coating “conforms to the shape and size of the cell aggregates”. Sufficient support for this amendment is found in the published application in paragraph 186 (present specification, paragraph 168). In response, the Examiner, in the Advisory Action mailed February 14, 2008, stated that the use of the term “conformal is not commensurate with that provided in the specification (p. 3 of the Advisory Action)”. Although Applicants respectfully disagree with this, Applicants present the following arguments which are commensurate with that described in the specification.

WO’159 describes a composition produced by two similar methods (Examples 1 and 3). Example 1 describes a composition whereby the “modified alginate, ionically cross-linked beads...were synthesized by conventional coaxial pneumatic nozzle technique (p. 35, lines 4-6 of WO’159)”. The alginate capsules are then exposed to photopolymerizing solution containing the photoinitiator. In Example 3, the composition is formed again by the coaxial nozzle which extrudes both the cells and the acrylated alginate PEG-diacrylate/photoinitiator solution, via the inner and outer nozzle, respectively. The extruded droplets fall into soybean oil forming a water-in-oil emulsion. In both Example 1 and 3, the photoinitiator is mixed with the photopolymerizable solution forming one solution, and upon light irradiation, activation and/or polymerization occurs throughout the solution (e.g., p. 39, lines 16-23 of WO’159). Because activation occurs throughout the one solution, and the size and shape of the capsule is pre-determined by the coaxial nozzle which forms the “spherical droplets”, the coating cannot be therefore said to be “conformal”.

In contrast, the claimed invention describes first putting the photoinitiator on the surface of the cell aggregate and then exposing the cells plus photoinitiator composition to the PEG photopolymerizing solution, which does not contain the photoinitiator. So, only the photoinitiator resides at the interface between the cells and the PEG solution. Thus, upon irradiation with light, the polymerization and coating formation can only occur at the surface of the cell aggregates, or interface between the cell and the PEG solution. Accordingly, the claimed method is also referred to as “interfacial photopolymerization (paragraphs 129 and 212)”. This interfacial photopolymerization method of conformally coating the cells is unique to the claimed invention and is not described in WO’159. WO’159 describes a combined solution having both

the photoinitiator and the alginate/PEG solution together, thus irradiation activates the entire solution.

A second Declaration by Dr. Xiaojie Yu, incorporated herein in its entirety, also speaks to the method producing the claimed composition which cannot be accomplished by the method described in WO'159. See Item 6, part (A).

The arguments provided herein are not inconsistent with those previously presented in the response filed December 21, 2007, which was not entered for allegedly containing new matter.

The claimed compositions have the advantage that they can encapsulate more cells per volume. Table 3 shows data for islets encapsulated by microcapsules, similar to the disclosure of WO'159. Table 4 shows data for islets encapsulated as per the compositions of the claimed invention. As discussed in the attached Yu Declaration (Item 6B), significantly higher cell densities may be achieved through conformal coatings. This means that for a given implant size, the number of islet cells that can be delivered is larger using conformal coatings according to the invention. This has positive implications for the potential implant recipient. For example, in the case of diabetes, by delivery of more islets/implant, the implant recipient can be free of the need for supplemental insulin injections.

The positive effect of the claimed invention is supported by the data in the present application. Using the conformally coated cells according to the claimed invention, implantation of both sub-human primate and human islets in mice provided regulation of blood glucose levels for extended periods (see Example 5 and Figures 7-14). Positive results were also obtained with *Cynomolgus* primates. Results are summarized in paragraph 0282 and Table 2 on page 64. Treatment of diabetic baboons is described in Example 7. For example, the first diabetic baboon implanted achieved insulin independence within 17 days post implant and continued without insulin through 180 days (paragraph 0287 & Figure 22). Although WO'159 discloses that their methods may be applied to cells, including islets (see Abstract, for example), no supporting data to support this assertion is provided.

In view of Applicants' amendments and arguments, reconsideration and withdrawal of the above ground of rejection is respectfully requested.

**No Disclaimers or Disavowals**

Although the present communication may include alterations to the application or claims, or characterizations of claim scope or referenced art, the Applicants are not conceding in this application that previously pending claims are not patentable over the cited references. Rather, any alterations or characterizations are being made to facilitate expeditious prosecution of this application. The Applicants reserve the right to pursue at a later date any previously pending or other broader or narrower claims that capture any subject matter supported by the present disclosure, including subject matter found to be specifically disclaimed herein or by any prior prosecution. Accordingly, reviewers of this or any parent, child or related prosecution history shall not reasonably infer that the Applicants have made any disclaimers or disavowals of any subject matter supported by the present application.

**Co-Pending Applications of Assignee**

Applicant to wishes to draw the Examiner's attention to the following co-pending applications of the present application's assignee. Entry in **Bold** is the present application.

Serial Number	Title	Filed
10/684859	<b>IMPLANTATION OF ENCAPSULATED BIOLOGICAL MATERIALS FOR TREATING DISEASES</b>	14-Oct-2003
11/037727	METHOD OF USING FIBRIN-BOUND ANGIOGENIC FACTORS TO STIMULATE VASCULARIZATION OF TRANSPLANT SITE OF ENCAPSULATED CELLS	18-Jan-2005
11/644606	GELS FOR ENCAPSULATION OF BIOLOGICAL MATERIALS	22-Dec-2006

**CONCLUSION**

In view of Applicants' amendments to the claims and the foregoing Remarks, it is respectfully submitted that the present application is in condition for allowance. Should the Examiner have any remaining concerns which might prevent the prompt allowance of the

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application, the Examiner is respectfully invited to contact the undersigned at the telephone number appearing below.

Please charge any additional fees, including any fees for additional extension of time, or credit overpayment to Deposit Account No. 11-1410.

Respectfully submitted,

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